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Research Article



The TGFβ Signaling Pathway Could Be Effective in the Pathogenesis of Human Lymphotropic Virus Type 1 Evidence from a Systems Biology Study

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Abstract

Background: Human T-cell lymphotropic virus type 1 (HTLV-1) is responsible for two critical human diseases: Adult T-cell leukemia/lymphoma (ATLL) and HTLV-1-associated myelopathy/tropical spastic paraparesis (HAM/TSP). Although most cases of HTLV-1 infection are asymptomatic and the prevalence of serious diseases like HAM/TSP and ATLL is not remarkable, their prognosis is poor. It has been assumed that various immunological responses, different interactions between the patient's body and the virus, and clearly the virus itself, are responsible for pathogenesis. To prevent and manage the consequences of this oncogenic virus, we should clearly understand the molecular pathways and interactions between the HTLV-1 virus and human cells.

Objectives: In this study, we aim to identify the most essential genes and microRNAs involved in the induction of ATLL disease and its laboratory confirmation.

Methods: After a purposeful search in databases and a qualitative review of obtained data, the data were processed using R software, and miRNAs with different expressions (DEMs) were calculated according to the value of logFC. The gene targets of miRNAs were obtained using the miRDB online tool. The protein-protein interaction network of gene targets was drawn and analyzed with the STRING database. The sub-network was extracted using Cytoscape. Pathway analysis was conducted, and proteins participating in the enriched pathways were selected to propose the implicated signaling network using the information available on the website ENRICHR and KEGG message transduction. An intracellular control gene, RPLPO, and three other key genes were selected and evaluated in 10 healthy participants, 10 asymptomatic carriers (ACs), and 10 ATLL patients using real-time PCR.

Results: Mir-20a-5p was identified as an essential microRNA with different expression in the patient group compared to the healthy and ACs. Transforming growth factor-beta 1 (TGF-β1), TGFBR1, and TGFBR2, which have a crucial role in disease pathogenesis, were identified.

Conclusions: The results demonstrated that TGFBRI's expression level was remarkably increased in ATLL patients compared to healthy donors and ACs. Further investigations are required to assess the prognostic and therapeutic function of these elements.

Keywords: Human T-Cell Lymphotrophic Virus Type 1, HTLV1, Adult T-Cell LeukemiA/Lymphoma, ATLL, miRNA, RTqPCR.

1. Background

Human T-cell lymphotropic virus type 1 (HTLV-1) is a member of the retrovirus family, which was first diagnosed in patients suffering from cutaneous T-cell lymphomas (1, 2). This virus has infected the human population for more than 20,000 years. This retrovirus can be transmitted through sex, from mother to child, and through blood transfusion. The virus involves lymphocytes, affects multiple organs through lymphocytes' movement, and causes various clinical manifestations (3). The Middle East is known today as an endemic region for HTLV-1 (4, 5). Although most patients with HTLV-1 infection have no clinical symptoms, the prevalence of severe diseases like adult T-cell leukemia/lymphoma (ATLL) and HTLV-1-associated myelopathy/tropical spastic paraparesis (HAM/TSP) is not remarkable, but their prognosis is poor. The mortality rate of ATLL patients is high because their treatment is very difficult. Therefore, a better understanding of HTLV-1 pathogenesis can lead to finding new ways for treatment (6, 7).

Bioinformatics is an interdisciplinary science that connects biological data with techniques for storing,

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distributing, and analyzing information to reinforce various fields of scientific experiments, such as biomedicine. Bioinformatics is fortified by highthroughput data-production investigations, such as the assessment of gene expression patterns and genomic sequence determinations (8, 9). Although the mechanisms of HTLV-1 are not yet fully understood, it has been assumed that various immunological responses, different interactions between the patient's body and the virus, and clearly the virus itself, are responsible for pathogenesis (10, 11). Evaluation of genetic background, cytokines, and pathways of HTLV-1 helps us to find better ways for diagnosis and treatment of this virus.

2. Objectives

To prevent and manage the consequences of this oncogenic virus, we should clearly understand the molecular pathways and interactions between the HTLV-1 virus and human cells. Since few studies evaluate the exact role of miRNAs and their targets in HTLV-1, this study aimed to run a high-throughput miRNA expression dataset to identify the key miRNAs and their target genes in HTLV-1 pathogenesis and find signaling pathways through which HTLV-1 affects the human body.

3. Methods

3.1. MicroRNA Expression Microarray Dataset and Different Expressions Analysis

The miRNA sample of CD4 T cells, including individual platforms with Accession number GSE31629, GPL 7731, was derived from the Gene Expression Omnibus (12) database. The dataset contains 62 separate samples of 40 ATLL patients and 22 healthy humans. Different expressions (DEMs) were identified between datasets. The first 48 miRNAs with higher expression variance values were selected. The DEMs were selected based on Benjamini-Hochberg FDR-adjusted P-values < 0.05.

3.2. Initial Pathway Analysis

DIANA-mirPath V3 was used to conduct the initial pathway analysis. The TarBase database V.7 and microT-CDS database V5 were used for pathway prediction. Heatmap plots from DIANA were generated using the pheatmap package in R 3.6.3. The common miRNA from heatmap results was obtained using a Venn diagram. A P-value < 0.05 was considered significant.

3.3. Predicting Genes Targeted by miRNAs and Constructing the Protein-Protein Interaction (PPI) Network

The miRDB online tool was used to find miRNA target genes. miRNA gene targets were sorted based on target rank. The target score for selected miRNA targets was obtained. The first gene symbols with target scores of 80 -100 were selected. To form protein-protein interactions, the STRING database was used. The PPI network of chosen DEM target genes was created using Cytoscapev3.8.0. The subnetwork was built in accordance with the first 267 proteins with higher degree values and betweenness. Nine modularities were obtained.

3.4. Functional Gene Enrichment and Proposal of Signaling Pathway

To perform gene analysis, the ENRICHR online tool was used. Genes related to the desired signaling pathway were selected based on enrichment results. The KEGG database was used to define cellular pathways and interactions among proteins.

3.5. Study Population and Sample Collection

This study was conducted on thirty participants [10 ATLL, 10 asymptomatic carriers (ACs), and 10 healthy individuals]. The ATLL patients were admitted to Shariati Hospital, Tehran, and Tehran province, Iran. The data for ACs and healthy participants were gathered from the Tehran Blood Transfusion Center. The inclusion criteria for healthy participants were no previous medication, no other diseases, no acute infection, no genetic disorders, and no history of vaccination during the last six months. The inclusion and exclusion criteria for ACs were similar to those for healthy participants, plus a positive PCR or ELISA test for HTLV-1. The ATLL patients were diagnosed based on the latest version of World Health Organization criteria and oncologist opinion. The healthy participants were chosen with negative ELISA and PCR tests and no clinical manifestations. They also must not have symptoms of acute infection and a history of vaccination during the last six months. Informed consent was obtained for experimentation with human subjects/samples.

3.6. RNA Extraction and cDNA Synthesis

Whole blood samples from 10 ATLL patients, 10 ACs, and 10 healthy individuals were acquired. Peripheral blood mononuclear cells (PBMCs) were isolated from EDTA-treated blood samples using a Ficoll density gradient. mRNAs were extracted from PBMCs using RNJia PB (ROJE Company). cDNAs were synthesized using random primers and reverse transcriptase based on RT-ROSET manufacturer guidelines (ROJE Company). To evaluate the accuracy of RNA extraction, RPLPO gene amplification was used, confirming the accuracy of both cDNA synthesis and RNA extraction.

3.7. Oligonucleotide Designing and Real-time PCR

A real-time PCR assay was performed to calculate RPLP0, transforming growth factor-beta 1 (TGF- β 1), TGF-BR1, and TGF-BR2 in both carrier and healthy groups, along with ATLL patients, using specific primers by Real-Time PCR in a Q 6000 Rotor-Gene machine (Qiagen, Germany). Table 1 displays the nucleotide sequence of primers. RPLP0 was relatively measured from 5-point standard curves. To calculate the normalized value of each gene expression, the Expression Index was used. The Expression Index is defined as the ratio of the relative copy number of the mRNA of interest to the relative mRNA copy number of the reference gene.

Targeted Genes	Sequence (5 $ ightarrow$ 3)
TGF-β	
Forward	GCCGACTACTACGCCAAGGA
Reverse	GAGCAACACGGGTTCAGGTAC
TGF-βR1	
Forward	GCAGAGCTGTGAAGCCTTGAG
Reverse	GCCTTCCTGTTGACTGAGTTGC
TGF-Br2	
Forward	AGGGCATCCAGATGGTGTGTGA
Reverse	GCTCCAGCTCACTGAAGCGTTC
HBZ	
Forward	ACGTCGCCCGGAGAAAACA
Reverse	CTCCACCTCGCCTTCCAACT
LTR	
Forward	GGCTCGCATCTCCCCTTCAC
Reverse	GAGCAAGCAGGGTCAGGCAA
Abbreviations: TGF-β	, transforming growth factor-beta.

3.8. Statistical Analysis

The experimental results are statistically presented as the mean \pm SD. Data analysis was carried out using Prism-GraphPad software. The Kolmogorov-Smirnov test was conducted to evaluate the normality of the variable distributions. The Kruskal-Wallis test was used to compare the data between the groups and to assess the correlation of the parameters. P-values lower than 0.05 were considered statistically significant.

4. Results

4.1. Different Expressions Analysis

Fifty miRNAs were finally selected, and the heatmap plot of their expression based on variances of expression was obtained. Ultimately, 49 DEMs with an absolute value of logFC greater than 2.5 were determined (Box 1).

miRNA ID	
hsa-miR-146b-5p	
hsa-miR-451	
hsa-miR-151-5p	
hsa-let-7b	
hsa-let-7c	
hsa-miR-29c	
hsa-miR-26a	
hsa-miR-342-3p	
hsa-miR-181a	
hsa-miR-29a	
hsa-miR-28-5p	
hsa-let-7g	
hsa-miR-92a	
hsa-miR-199a-3p	
hsa-miR-342-5p	
hsa-miR-30b	
hsa-miR-221	
hsa-miR-320a-5p	
hsa-miR-361-5p	
hsa-miR-423-5p	
hsa-miR-20a	
hsa-miR-17	
hsa-miR-10A	
hsa-miR-125B	
hsa-miR-212	
hsa-miR-140-3p	
hsa-miR-20b	
hsa-let-7a	
hsa-miR-186	
hsa-miR-148b	
hsa-miR-361-3p	
hsa-miR-150	
hsa-miR-26b	
hsa-miR-101	
hsa-miR-103	
hsa-miR-30d	
hsa-miR-30c	
hsa-miR-24	
hsa-miR-27a	
hsa-miR-374a	
hsa-let-7f	
hsa-miR-19b	
hsa-miR-494	
hsa-miR-324-3p	
hsa-miR-23b	
hsa-miR-98	
hsa-miR-142-3P	
hsa-miR-210	
hea-miP-004	

4.2. Initial Pathway Analysis

To identify biologically significant miRNAs, the relationship between DEMs and biological pathways was explored. The data were analyzed in DIANA based on TarBase and microTCDS. The heatmap results were obtained, and mir-20a-5p was found to be related to all

Table 2. The Combined Score of Each Signaling Pathway				
Term	Combined Score			
Axon guidance	78.98587			
MAPK signaling pathway	41.74268			
Circadian rhythm	117.4687			
Endocytosis	26.76269			
Autophagy	26.86101			
TGF-beta signaling pathway	29.91466			
RNA degradation	16.61342			
Cholesterol metabolism	19.22018			
Gastric acid secretion	2.09712			



Figure 1. A, Protein-protein interaction network of different expressions (DEMs) in human T-cell lymphotropic virus type 1 (HTLV-1) infected cases; B, modularity 1: Endocytosis; C, modularity 2: Circadian rhythm pathway; D, modularity 3: Autophagy pathway; E, modularity 4: Axon guidance pathway; F, modularity 5: Transforming growth factor-beta (TGFβ) pathway; G, modularity 6: Cholesterol metabolism; H, modularity 7: Gastric acid secretion pathway; I, modularity 8: RNA degeneration pathway; J, modularity 9: MAPK signaling pathway.

desired pathways. The target genes of mir-20a-5p, along with their scores and target ranks, were calculated, and 685 genes with target scores of 80 - 100 were selected using ENRICHR. The combined score of each signaling pathway was calculated and presented in Table 2.

Desired signaling pathways were chosen using both Heatmaps and ENRICHR.

4.3. Construction of the PPI Network and Extraction of the PPI Sub-network

The protein-protein interaction network was used to demonstrate the strong relationships between DEMs. This network contains 266 nodes and 1134 edges (Appendix 1 in Supplementary File). The size of the nodes represents the degree of each node, and the color of each node indicates the level of expression of each protein. Based on Cytoscape results, the sub-networks were obtained. Nine modularities were identified: Modularity 1 is the endocytosis pathway, modularity 2 is the circadian rhythm pathway, modularity 3 is the autophagy pathway, modularity 4 is the axon guidance pathway, modularity 5 is the transforming growth factor-beta (TGF- β) pathway, modularity 6 is cholesterol metabolism, modularity 7 is the gastric acid secretion pathway, modularity 8 is the RNA degeneration pathway, and modularity 9 is the MAPK signaling pathway (Figure 1A - J).

4.4. Functional Gene Enrichment and Proposal of Signaling Network

Mir-20a-5p targets the TGF- β signaling pathway. The TGF- β signaling pathway was identified by both heatmap plots and ENRICHR. According to enrichment results, genes and proteins related to the TGF- β signaling pathway were determined. SMAD4, BMPR2, RBL1, ZFYVE9, BAMBI, MAPK1, RGMB, E2F5, SMAD5, RGMA, and TGFBR2 are the genes involved in the TGF- β enriched signaling pathway.

4.5. Descriptive Analysis of Patients

The study evaluated host-microbe interactions using gene expression measurements in 10 healthy participants, 10 ACs, and 10 ATLL patients. The mean age in the ATLL group was 53.4 ± 12.98 , in the ACs was 53.9 ± 6.641 , and in the healthy group was 57.2 ± 2.936 . There was no significant relationship between groups according to age (P = 0.43). In the ATLL group, 20% were females and 80% were males, while in both the ACs and healthy groups, all participants were males. Therefore, there were significant differences in gender between groups (P < 0.05) (Table 3).

Table	3.	Demographic	Characteristics	(Gender	and	Age	of	Adult	T-Cel
Leukemia/Lymphoma Patients, ACs and Healthy Donors) ^a									

Variables	Normal	ACs	ATLL	P-Value
Age	57.2 ± 2.936	53.9 ± 6.641	53.4 ± 12.98	0.43
Gender				0.05
Female	0	0	2(20)	
Male	10 (100)	10 (100)	8 (80)	

Abbreviations:	ACs,	asymptomatic	carriers;	ATLL,	adult	T-cell
leukemia/lymphoma.						

^a Values are expressed as mean ± SD or No. (%).

4.6. Expression Level of Transforming Growth Factor-Beta 1 Among Healthy, Asymptomatic Carriers, and Adult T-Cell Leukemia/Lymphoma

The expression level of TGF- β 1 in ATLL patients was lower than in the ACs and healthy group, but it was not clinically significant (P = 0.5092). Additionally, the expression level of TGF- β 1 was higher in healthy participants compared to ACs (Figure 2A).

4.7. Expression Level of TGFBR1 Among Healthy, Asymptomatic Carriers, and Adult T-Cell Leukemia/Lymphoma

The expression level of TGFBR1 in ATLL patients was higher than in the ACs and healthy group, and this difference was clinically significant (P = 0.0011). Additionally, the expression level of TGFBR1 was higher in ACs compared to healthy participants (Figure 2B).

4.8. Expression Level of TGFBR2 Among Healthy, Asymptomatic Carriers, and Adult T-Cell Leukemia/Lymphoma

The expression level of TGFBR2 in ATLL patients and ACs was lower than in the healthy group, and this difference was clinically significant (P = 0.0436). Furthermore, the expression level of TGFBR2 was lower in ACs compared to healthy participants (Figure 2C).

5. Discussion

One of the most important results of our study was the role of TGFBR1 in ATLL. The TGFBR1 gene commands the synthesis of TGF- β receptor type 1 protein. The function of this receptor is to transmit signals from the cell surface into the cell via the signal transduction process (13, 14). Since TGFBR1 prevents cells from growing and dividing too rapidly, it can function as a tumor suppressor (15). We found that TGFBR1 gene expression is significantly higher in ATLL patients than in ACs and healthy donors. However, contrary to our results, some studies have demonstrated that decreased TGFBR1 signaling may contribute to cancer progression (16, 17). We hypothesize that TGFBR1 expression increases to defend against cancer cells but cannot overcome them, resulting in higher gene expression than in healthy donors. Another possible explanation is the difference in the type of cancer.

Figure 2. A, Expression level of transforming growth factor-beta 1 (TGF-β1) among healthy, asymptomatic carriers (ACs) and adult T-cell leukemia/lymphoma (ATLL); B, expression level of TGFBR1 among healthy, ACs and ATLL

The TGFBR2 gene encodes TGF- β receptor type 2, which is part of the serine/threonine protein kinase family. TGFBR2 is a transmembrane protein receptor with a protein kinase domain that forms a heterodimeric complex with another receptor protein and links TGF-beta. This receptor/ligand complex regulates cell proliferation through protein phosphorylation (18, 19). Mutations in this gene are associated with various diseases, including the development of different types of tumors (20). We found that the expression level of TGFBR2 in ATLL patients and ACs was significantly lower than in healthy donors. Our results suggest that TGFBR2 acts as a tumor suppressor. Consistent with our study, several studies have shown that a decrease in TGFBR2 expression level is associated with higher grades of cancer (21-23). We can conclude that in the early stages of cancer, TGFBR2 acts as a tumor suppressor.

One of the vital signaling networks is the transforming growth factor TGF- β signaling pathway, which plays an essential role in cell proliferation, inflammation, apoptosis, angiogenesis, and tumor genesis (24). Although the level of TGF- β is nearly the same in ATLL patients and healthy humans (25), *in vitro* studies show that in ATLL patients, the expression of

TGF- β mRNA and secretion of TGF- β is higher than in healthy humans (26-28). While TGF- β usually inhibits cell proliferation (29), it has no inhibitory effect on the proliferation of T-cells infected with HTLV-1 (30).

Transforming growth factor-beta (TGFβ) is a pleiotropic multifunctional cytokine secreted from epithelial cells and fibroblasts (31, 32). When TGF- β family ligands bind to the active TGFB type II serine/threonine kinase receptor (TBRII), they recruit the type I receptor (T β RI) and form a stable oligometric receptor complex. The formation of this complex causes the type II receptor to phosphorylate the type I receptor. This phosphorylation activates the type I receptorkinase through structural changes. SMAD2/3 phosphorylation results in binding to SMAD4 and the creation of a signaling complex, which translocates to the nucleus to induce downstream changes in gene expression through the canonical signaling pathway.

After nuclear translocation, to regulate gene expression, the SMAD complex binds to other transcription factors or chromatin-remodeling proteins. Transforming growth factor-beta 1 also functions through a plethora of noncanonical SMAD-independent pathways, including the mitogen-activated protein (MAP) kinase pathway, which signals via extracellular

Brieflands

signal-regulated kinase 1/2 or TGF- β -activated kinase 1 (TAK1) (33, 34). Several studies have demonstrated that TGF- β is deregulated in cancers and plays an essential role in tumor initiation and progression. In the early stage of tumors, TGF- β acts as a tumor suppressor through its antiproliferative effects, while in the later stages, it acts as a tumor promoter, aiding tumor metastasis through an autocrine TGF- β loop (35, 36).

Among the three TGF- β isoforms, TGF- β 1 is the most essential, controlling differentiation, hemostasis, and the development of other immune cells. Transforming growth factor-beta 1 is a potent inhibitor of cell proliferation in most endothelial, epithelial, and hematopoietic cells, such as T lymphocytes. Transforming growth factor-beta 1 suppresses tumor cells in their early stages. However, tumor cells develop mechanisms to escape the inhibitory effects of TGF- β 1. Some mutations occur in TGF- β receptors, for example, downstream in TGFBR1 and TGFBR2, SMAD2, and SMAD4, which occur in metastatic colon cancer and pancreatic cancer (37).

Kim et al. found that in ATL cells, TGF- β is overproduced, and the main isoform secreted by these cells is TGF- β 1. Cytokines such as IL-10 and TGF- β suppress the immune system in ATLL patients. Although the TGF- β level in the serum of ATLL patients is not significantly higher than in healthy people, *in vitro* studies of ATLL cell culture and long-term ATLL T-cell lines have proven that ATLL cells express TGF- β mRNA and secrete TGF- β cytokines. Although TGF- β is a wellknown cell proliferation inhibitor, it cannot prevent the spontaneous proliferation of HTLV-1-infected T cells. Kim et al. found that the expression of TGF- β 1 in leukemic cells of ATLL patients and HTLV-1-infected T-cells is high (28).

Following our study, Javle et al. found that the TGF- β signaling pathway plays an important role in pancreatic cancer. They demonstrated that the TGF- β 1 plasma level was high in pancreatic cancer; however, the expression level of TGFBR2 and SMAD4 was not related to pancreatic cancer (38). Hadi et al. showed that TGFBR1 and TGFBR2 have crucial roles in chronic lymphocytic leukemia (CLL), and five deregulated miRNAs, including mir-574, mir-499, mir-125b, mir-106a, and mir-9, are involved in CLL pathogenesis (39).

Mir-20a-5p is a component of the mir-17-92 cluster, which is associated with many kinds of cancers. Overexpression or downregulation of miR-20a-5p participates in downstream signaling pathways, such as PI3K-Akt, MAPK, and TGF- β signaling pathways.

Controlling incessant proliferation signals, evading growth inhibition, activating invasion and metastasis, enabling limitless replication, promoting angiogenesis, resisting cellular death, and avoiding immune destruction are mechanisms of miR-20a-5p (12). Wei Huang et al. have demonstrated that deregulated miRNAs are one of the vital causes of cancers, making them biomarkers for cancer diagnosis and prognosis (40).

In this study, we identified the TGF- β signaling pathway as the most important signaling pathway in ATLL pathogenesis. Mohadeseh Valizadeh et al. demonstrated that downregulation in the TGF-B signaling pathway is associated with sulfur mustard (SM) skin lesions. They showed that TGF-β, TGFBR2, and their functional miRNAs, including mir-20a and mir-21a, contribute to SM lesions (41). Zhang et al. determined that mir-20a is an independent prognostic and therapeutic factor for colorectal cancer (42). In this study, we also demonstrated that SMAD4, BMPR2, RBL1, ZFYVE9, BAMBI, MAPK1, RGMB, E2F5, SMAD5, RGMA, and TGFBR2 genes are involved in the TGF-β signaling pathway. TGF-beta family members, which include TGFbetas, bone morphogenetic proteins (BMPs), and activins, are structurally related secreted cytokines with a broad spectrum of cellular functions (43, 44).

5.1. Conclusions

In this study, which resulted from the bioinformatic analysis of high-throughput microarray data, it was determined that TGFBR1 has a more active role than TGFBR2 and even TGF-beta itself. Additionally, Mir-20a-5p and the genes SMAD4, BMPR2, RBL1, ZFYVE9, BAMBI, MAPK1, RGMB, E2F5, SMAD5, RGMA, and TGFBR2 were identified as main genes in the TGF-beta signaling pathway. More studies are needed to further clarify the role of TGFBR1.

Supplementary Material

Supplementary material(s) is available here [To read supplementary materials, please refer to the journal website and open PDF/HTML].

Footnotes

Authors' Contribution: Writing-review & editing, writing-original draft, project administration, methodology, investigation, formal analysis: Z. A., M. M., A. B., and A. A.; Conceptualization, data curation,

methodology, writing-original draft: N. T.; Writingreview & editing, writing-original draft, validation, supervision, data curation, conceptualization: L. L.; Writing-review & editing, writing-original draft, supervision, data curation, conceptualization: S. M.

Conflict of Interests Statement: The authors declare no conflict of interest.

Data Availability: The dataset presented in the study is available on request from the corresponding author during submission or after publication.

Ethical Approval: This work has been carried out in accordance with the ethics of the World Medical Association (Declaration of Helsinki) for experiments involving humans. Also, the study was approved by the Ethics Committee of Kerman University of Medical Sciences, Kerman, Iran (ethics code: IR.KMU.AH.REC.1401.110).

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References

- Poiesz BJ, Ruscetti FW, Gazdar AF, Bunn PA, Minna JD, Gallo RC. Detection and isolation of type C retrovirus particles from fresh and cultured lymphocytes of a patient with cutaneous T-cell lymphoma. *Proc Natl Acad Sci U S A*. 1980;77(12):7415-9. [PubMed ID: 6261256]. [PubMed Central ID: PMC350514]. https://doi.org/10.1073/pnas.77.12.7415.
- Poiesz BJ, Ruscetti FW, Reitz MS, Kalyanaraman VS, Gallo RC. Isolation of a new type C retrovirus (HTLV) in primary uncultured cells of a patient with Sezary T-cell leukaemia. *Nature*. 1981;**294**(5838):268-71. [PubMed ID: 6272125]. https://doi.org/10.1038/294268a0.
- Martinez MP, Al-Saleem J, Green PL. Comparative virology of HTLV-1 and HTLV-2. *Retrovirology*. 2019;16(1):21. [PubMed ID: 31391116]. [PubMed Central ID: PMC6686503]. https://doi.org/10.1186/s12977-019-0483-0.
- Proietti FA, Carneiro-Proietti AB, Catalan-Soares BC, Murphy EL. Global epidemiology of HTLV-I infection and associated diseases. Oncogene. 2005;24(39):6058-68. [PubMed ID: 16155612]. https://doi.org/10.1038/sj.onc.1208968.
- Yoshida M, Miyoshi I, Hinuma Y. Isolation and characterization of retrovirus from cell lines of human adult T-cell leukemia and its implication in the disease. *Proc Natl Acad Sci U S A*. 1982;**79**(6):2031-5. [PubMed ID: 6979048]. [PubMed Central ID: PMC346116]. https://doi.org/10.1073/pnas.79.6.2031.
- Ishitsuka K, Tamura K. Human T-cell leukaemia virus type I and adult T-cell leukaemia-lymphoma. *Lancet Oncol.* 2014;**15**(11):e517-26. [PubMed ID: 25281470]. https://doi.org/10.1016/S1470-2045(14)70202-5.

- Manns A, Hisada M, La Grenade L. Human T-lymphotropic virus type I infection. *Lancet.* 1999;**353**(9168):1951-8. [PubMed ID: 10371587]. https://doi.org/10.1016/s0140-6736(98)09460-4.
- Barnes MR. Bioinformatics for geneticists: A bioinformatics primer for the analysis of genetic data. Hoboken, New Jersey: John Wiley & Sons; 2007.
- 9. Jones NC, Pevzner PA. *An introduction to bioinformatics algorithms*. Cambridge, Massachusetts: MIT Press; 2004.
- Rizkallah G, Alais S, Futsch N, Tanaka Y, Journo C, Mahieux R, et al. Dendritic cell maturation, but not type I interferon exposure, restricts infection by HTLV-1, and viral transmission to T-cells. *PLoS Pathog.* 2017;13(4). e1006353. [PubMed ID: 28426803]. [PubMed Central ID: PMC5413061]. https://doi.org/10.1371/journal.ppat.1006353.
- Furuta R, Yasunaga JI, Miura M, Sugata K, Saito A, Akari H, et al. Human T-cell leukemia virus type 1 infects multiple lineage hematopoietic cells in vivo. *PLoS Pathog.* 2017;**13**(11). e1006722. [PubMed ID: 29186194]. [PubMed Central ID: PMC5724899]. https://doi.org/10.1371/journal.ppat.1006722.
- Papageorgiou SG, Kontos CK, Tsiakanikas P, Stavroulaki G, Bouchla A, Vasilatou D, et al. Elevated miR-20b-5p expression in peripheral blood mononuclear cells: A novel, independent molecular biomarker of favorable prognosis in chronic lymphocytic leukemia. *Leuk Res.* 2018;**70**:1-7. [PubMed ID: 29715621]. https://doi.org/10.1016/j.leukres.2018.04.014.
- 13. Pasche B, Luo Y, Rao PH, Nimer SD, Dmitrovsky E, Caron P, et al. Type I transforming growth factor beta receptor maps to 9q22 and exhibits a polymorphism and a rare variant within a polyalanine tract. *Cancer Res.* 1998;**58**(13):2727-32. [PubMed ID: 9661882].
- Pasche B, Kolachana P, Nafa K, Satagopan J, Chen Y, Lo RS, et al. TβR-I (6A) is a candidate tumor susceptibility allele. *Cancer Res.* 1999;**59**(22):5678-82.
- Pasche B, Pennison MJ, Jimenez H, Wang M. TGFBR1 and cancer susceptibility. *Transac Am Clin Climatol Assoc.* 2014;125:300.
- Chen T, de Vries EGE, Hollema H, Yegen HA, Vellucci VF, Strickler HD, et al. Structural alterations of transforming growth factor-? receptor genes in human cervical carcinoma. *Int J Cancer*. 1999;**82**(1):43-51. https://doi.org/10.1002/(sici)1097-0215(19990702)82:1<43::Aidijc9>3.0.Co;2-0.
- Pasche B. Role of transforming growth factor beta in cancer. *J Cell Physiol.* 2001;**186**(2):153-68. [PubMed ID: 11169452]. https://doi.org/10.1002/1097-4652(200002)186:2<153::AID-JCP1016>3.0.CO;2-J.
- Simcikova D, Heneberg P. Refinement of evolutionary medicine predictions based on clinical evidence for the manifestations of Mendelian diseases. *Sci Rep.* 2019;9(1):18577. [PubMed ID: 31819097]. [PubMed Central ID: PMC6901466]. https://doi.org/10.1038/s41598-019-54976-4.
- Guerrero-Esteo M, Sanchez-Elsner T, Letamendia A, Bernabeu C. Extracellular and cytoplasmic domains of endoglin interact with the transforming growth factor-beta receptors I and II. *J Biol Chem.* 2002;**277**(32):29197-209. [PubMed ID: 12015308]. https://doi.org/10.1074/jbc.M111991200.
- Singh KK, Rommel K, Mishra A, Karck M, Haverich A, Schmidtke J, et al. TGFBR1 and TGFBR2 mutations in patients with features of Marfan syndrome and Loeys-Dietz syndrome. *Hum Mutat*. 2006;**27**(8):770-7. [PubMed ID: 16799921]. https://doi.org/10.1002/humu.20354.
- 21. Gobbi H, Arteaga CL, Jensen RA, Simpson JF, Dupont WD, Olson SJ, et al. Loss of expression of transforming growth factor beta type II receptor correlates with high tumour grade in human breast in-situ and invasive carcinomas. *Histopathology*. 2000;**36**(2):168-77. [PubMed ID: 10672063]. https://doi.org/10.1046/j.1365-2559.2000.00841.x.

- Figueroa JD, Flanders KC, Garcia-Closas M, Anderson WF, Yang XR, Matsuno RK, et al. Expression of TGF-beta signaling factors in invasive breast cancers: relationships with age at diagnosis and tumor characteristics. *Breast Cancer Res Treat.* 2010;**121**(3):727-35. [PubMed ID: 19937272]. [PubMed Central ID: PMC4159718]. https://doi.org/10.1007/s10549-009-0590-z.
- 23. Buck MB, Fritz P, Dippon J, Zugmaier G, Knabbe C. Prognostic significance of transforming growth factor beta receptor II in estrogen receptor-negative breast cancer patients. *Clin Cancer Res.* 2004;**10**(2):491-8. [PubMed ID: 14760070]. https://doi.org/10.1158/1078-0432.ccr-0320-03.
- Fabregat I, Fernando J, Mainez J, Sancho P. TGF-beta signaling in cancer treatment. *Curr Pharm Des.* 2014;20(17):2934-47. [PubMed ID: 23944366]. https://doi.org/10.2174/13816128113199990591.
- Inagaki A, Ishida T, Ishii T, Komatsu H, Iida S, Ding J, et al. Clinical significance of serum Th1-, Th2- and regulatory T cells-associated cytokines in adult T-cell leukemia/lymphoma: High interleukin-5 and -10 levels are significant unfavorable prognostic factors. *Int J Cancer*. 2006;**118**(12):3054-61. [PubMed ID: 16425276]. https://doi.org/10.1002/ijc.21688.
- Tendler CL, Greenberg SJ, Burton JD, Danielpour D, Kim SJ, Blattner WA, et al. Cytokine induction in HTLV-I associated myelopathy and adult T-cell leukemia: Alternate molecular mechanisms underlying retroviral pathogenesis. *J Cell Biochem*. 1991;46(4):302-11. [PubMed ID: 1757474]. https://doi.org/10.1002/jcb.240460405.
- Niitsu Y, Urushizaki Y, Koshida Y, Terui K, Mahara K, Kohgo Y, et al. Expression of TGF-beta gene in adult T cell leukemia. *Blood.* 1988;71(1):263-6. [PubMed ID: 2891388].
- 28. Kim SJ, Kehrl JH, Burton J, Tendler CL, Jeang KT, Danielpour D, et al. Transactivation of the transforming growth factor beta 1 (TGF-beta 1) gene by human T lymphotropic virus type 1 tax: a potential mechanism for the increased production of TGF-beta 1 in adult T cell leukemia. *J Exp Med.* 1990;**172**(1):121-9. [PubMed ID: 2358774]. [PubMed Central ID: PMC2188182]. https://doi.org/10.1084/jem.172.1.121.
- Huang SS, Huang JS. TGF-beta control of cell proliferation. J Cell Biochem. 2005;96(3):447-62. [PubMed ID: 16088940]. https://doi.org/10.1002/jcb.20558.
- Höllsberg P, Ausubel LJ, Hafler DA. Human T cell lymphotropic virus type I-induced T cell activation. Resistance to TGF-beta 1-induced suppression. J Immunol. 1994;153(2):566-73. https://doi.org/10.4049/jimmunol.153.2.566.
- Roberts AB, Heine UI, Flanders KC, Sporn MB. Transforming growth factor-beta. Major role in regulation of extracellular matrix. *Ann N Y Acad Sci.* 1990;**580**:225-32. [PubMed ID: 2186691]. https://doi.org/10.1111/j.1749-6632.1990.tb17931.x.
- Massague J. TGFbeta signalling in context. Nat Rev Mol Cell Biol. 2012;13(10):616-30. [PubMed ID: 22992590]. [PubMed Central ID: PMC4027049]. https://doi.org/10.1038/nrm3434.

- Wrana JL, Attisano L, Wieser R, Ventura F, Massague J. Mechanism of activation of the TGF-beta receptor. *Nature*. 1994;**370**(6488):341-7. [PubMed ID: 8047140]. https://doi.org/10.1038/370341a0.
- Yamashita H, ten Dijke P, Franzen P, Miyazono K, Heldin CH. Formation of hetero-oligomeric complexes of type I and type II receptors for transforming growth factor-beta. *J Biol Chem.* 1994;269(31):20172-8. [PubMed ID: 8051105].
- Dahmani A, Delisle JS. TGF-beta in T Cell Biology: Implications for Cancer Immunotherapy. Cancers (Basel). 2018;10(6). [PubMed ID: 29891791]. [PubMed Central ID: PMC6025055]. https://doi.org/10.3390/cancers10060194.
- Chaudhury A, Howe PH. The tale of transforming growth factor-beta (TGFbeta) signaling: A soigne enigma. *IUBMB Life*. 2009;61(10):929-39.
 [PubMed ID: 19787707]. [PubMed Central ID: PMC2810629]. https://doi.org/10.1002/iub.239.
- Kaklamani VG, Pasche B. Role of TGF-beta in cancer and the potential for therapy and prevention. *Expert Rev Anticancer Ther.* 2004;4(4):649-61. [PubMed ID: 15270668]. https://doi.org/10.1586/14737140.4.4.649.
- Javle M, Li Y, Tan D, Dong X, Chang P, Kar S, et al. Biomarkers of TGFbeta signaling pathway and prognosis of pancreatic cancer. *PLoS One*. 2014;9(1). e85942. [PubMed ID: 24465802]. [PubMed Central ID: PMC3896410]. https://doi.org/10.1371/journal.pone.0085942.
- Hadi N, Namazi F, Ketabchi F, Khosravian F, Nateghi B, Talebi A, et al. miR-574, miR-499, miR-125b, miR-106a, and miR-9 potentially target TGFBR-1 and TGFBR-2 genes involving in inflammatory response pathway: Potential novel biomarkers for chronic lymphocytic leukemia. *Pathol Res Pract.* 2022;**238**:154077. [PubMed ID: 36037658]. https://doi.org/10.1016/j.prp.2022.154077.
- Huang W, Wu Y, Qiao M, Xie Z, Cen X, Huang X, et al. CircRNA-miRNA networks in regulating bone disease. *J Cell Physiol*. 2022;237(2):1225-44. [PubMed ID: 34796958]. https://doi.org/10.1002/jcp.30625.
- Valizadeh M, Mirzaei B, Tavallaei M, Noorani MR, Amiri M, Soroush MR, et al. Down-regulation of TGF-b1, TGF-b receptor 2, and TGF-bassociated microRNAs, miR-20a and miR-21, in skin lesions of sulfur mustard-exposed Iranian war veterans. *J Recept Signal Transduct Res.* 2015;35(6):634-9. [PubMed ID: 26498464]. https://doi.org/10.3109/10799893.2015.1041646.
- Zhang G, Li YU, Zhou HE, Xiao H, Zhou T. miR-20a is an independent prognostic factor in colorectal cancer and is involved in cell metastasis. *Mol Med Rep.* 2014;10(1):283-91. https://doi.org/10.3892/mmr.2014.2144.
- Tzavlaki K, Moustakas A. TGF-beta Signaling. *Biomolecules*. 2020;10(3). [PubMed ID: 32210029]. [PubMed Central ID: PMC7175140]. https://doi.org/10.3390/biom10030487.
- Miyazono K. TGF-beta signaling by Smad proteins. Cytokine Growth Factor Rev. 2000;11(1-2):15-22. [PubMed ID: 10708949]. https://doi.org/10.1016/s1359-6101(99)00025-8.