



Pomegranate Seed Oil Boosts Memory by Reducing Amyloid Plaques and Restoring Neuronal Density in Alzheimer's Model Rats

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Abstract

Background: Pomegranate seed oil possesses antioxidant, anti-inflammatory, and neuroprotective properties. Alzheimer's disease is a prevalent neurodegenerative disorder that results in behavioral and cognitive impairments.

Objectives: This study aimed to investigate the effects of pomegranate seed oil on memory impairment, neuronal density, and amyloid beta (A β) plaque in the hippocampus of rats, both pre- and post-scopolamine administration.

Methods: Fifty-six male Wistar rats were randomly assigned to seven groups. Rats received intraperitoneal injections of pomegranate seed oil (0.32 and 0.64 mg/kg) or saline for 14 days, either before or after intraperitoneal infusion of scopolamine (3 mg/kg). Memory function was assessed using the passive avoidance test. Cresyl violet and Congo red staining techniques were employed to evaluate hippocampal neuronal density and A β plaques, respectively.

Results: Evaluation of memory using the passive avoidance test after scopolamine injection revealed a significant reduction in memory retention. Treatment with pomegranate seed oil at a dose of 0.32 mg/kg improved memory performance both before and after scopolamine injection; however, this improvement was not statistically significant. Scopolamine significantly increased the number of amyloid plaques ($P < 0.0001$) and significantly decreased the number of neurons in the hippocampus ($P < 0.0001$) compared to control rats. Pomegranate seed oil at doses of 0.32 and 0.64 mg/kg significantly reduced the density of amyloid plaques ($P < 0.0001$) and increased the number of neurons in the hippocampus when compared to saline-treated rats.

Conclusions: Pomegranate seed oil demonstrated significant neuroprotective and therapeutic effects by enhancing neuronal density and reducing amyloid plaque formation in the hippocampus. Therefore, pomegranate seed oil is suggested as a potential treatment for alleviating symptoms associated with Alzheimer's disease.

Keywords: Alzheimer's Disease, Amyloid Plaques, Hippocampus, Memory, Pomegranate Seed Oil, Scopolamine

1. Background

Alzheimer's disease (AD) and other forms of dementia rank as the sixth leading cause of death among older adults globally. The AD induces neurodegeneration, impairing various cognitive functions such as attention, language, orientation, and judgment. The primary symptoms include progressive memory loss and cognitive disorder, ultimately resulting in death (1). Key features of AD include

disrupted synapses, accumulated amyloid beta (A β) plaques, neurofibrillary tangles, and neuronal loss (2). Neuronal reduction is influenced by degenerative processes like A β plaques, oxidative stress, and inflammation (3). The cortex and hippocampus are particularly vulnerable to these degenerations early in the disorder. The hippocampus, crucial for learning and memory, undergoes atrophy due to AD-related pathological mechanisms (4). Despite extensive research, many FDA-approved AD drugs are ineffective,

unable to cure or slow AD progression, costly, and often cause adverse side effects (5). Consequently, researchers are exploring alternative products to mitigate AD pathogenesis, aiming for more accessible treatments with fewer side effects.

Recent studies highlight the significant role of oxidative and inflammatory processes in AD pathogenesis, contributing to senile plaques and neurofibrillary tangles. The abnormal accumulation of A β and tau proteins exacerbates redox imbalance, leading to oxidative stress associated with A β or tau-induced neurotoxicity. Evidence suggests oxidative stress enhances A β aggregation and facilitates tau protein polymerization and phosphorylation, initiating a negative cycle that contributes to AD onset and progression. Therefore, developing therapeutic strategies targeting neuroinflammation and oxidative stress is crucial in combating AD pathogenesis (6).

Natural products are vital sources for drug discovery, with compounds derived from them extensively evaluated for developing novel AD treatments (5). Pomegranate (*Punica granatum* L.), cultivated in West Asia, the Mediterranean, and other regions, contains compounds like hydrolysable and condensed tannins, flavanols, anthocyanins, phenolics, and natural acids, studied for their health benefits. Compared to other fruits and vegetables, pomegranates have higher concentrations of antioxidant polyphenolic compounds, providing neuroprotective benefits in various models (7). Pomegranate seed oil (PSO) is rich in linolenic, linoleic, punicic, stearic, palmitic, and oleic acids (8). Punicic acid exhibits antioxidant, anti-inflammatory, nephroprotective, hepatoprotective, neuroprotective, anti-cancer properties, improves carbohydrate metabolism, and reduces insulin resistance (9). The PSO improves sensorimotor function and memory performance dose-dependently (10). It acts as an antioxidant by scavenging reactive oxygen species (ROS) and restoring cell viability (11). Phenolic compounds in PSO show neuroprotective effects, reducing A β -induced toxicity, demonstrating antioxidant activity, and protecting against A β -induced cell death (12). Neuronal degeneration is controlled by inhibiting inflammation and amyloidogenesis in interleukin-1 β treated SK-N-SH cells (13). Pomegranates reduce soluble A β levels and A β deposition as plaques, improving behavioral performance in APPsw/Tg2576 mice (14). Pomegranate seed extract ameliorates memory impairments following bilateral common carotid artery occlusion (15). Pomegranate extract exhibits significant bioactive properties, including anti-inflammatory and antioxidant effects (6). Thus,

pathways regulating neuroinflammation and oxidative stress contribute to pomegranate's neuroprotective effects.

Scopolamine is widely used in neuroscience research to induce cognitive disorders in experimental models due to its ability to cross the blood-brain barrier. In AD, it is significant as it triggers cholinergic dysfunction and enhances A β deposition – key disease features (16). Scopolamine administration induces oxidative stress-mediated activation of the c-Jun N-terminal kinase (JNK) pathway, leading to synaptic dysfunction, neuroinflammation, and neurodegeneration. It induces lipid peroxidation (LPO) and ROS production while downregulating antioxidant proteins like Nrf2 and HO-1. Scopolamine promotes neuronal loss by upregulating pro-apoptotic markers such as Bax, Pro-Caspase-3, and Caspase-3 while decreasing the anti-apoptotic protein Bcl-2 (17). Despite numerous studies on pomegranate, further investigations are needed to understand pomegranate-related cellular and molecular mechanisms in animal models to confirm its efficacy as a therapeutic factor (18).

2. Objectives

This study aimed to evaluate the effects of PSO on memory, hippocampal neuronal density, and amyloid plaques in a scopolamine-induced AD model in rats.

3. Methods

3.1. Animals

The experimental protocol adhered to the National Institutes of Health Guide for the Care and Use of Laboratory Animals (NIH Publications No. 8023, revised 1978). The study received approval from the Research Ethics Committee of Golestan University of Medical Sciences, Gorgan, Iran (Agreement License No. IR.GOUMS.REC.1397.260). In this study, 56 male Wistar rats, weighing between 180 and 220 g, were obtained from the animal facility of Golestan University of Medical Sciences in Gorgan, Iran. Optimal conditions were maintained at a temperature of 22 ± 3 °C with a 12-hour light/dark cycle, and the rats had free access to standard food and drinking water.

3.2. Experimental Design

The rats were randomly assigned to seven groups (n = 8 per group). All animals underwent behavioral testing and received daily treatments as follows:

- Control group: Received no injection.

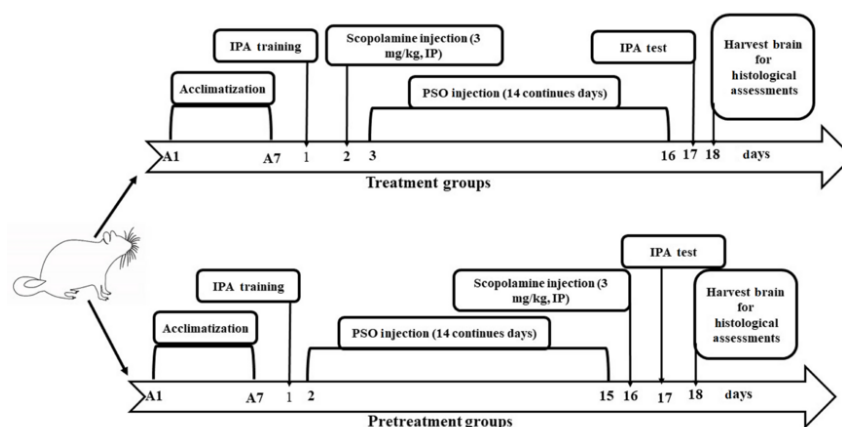


Figure 1. Schematic representation of the experimental procedure. Abbreviations: IPA, Inhibitory passive avoidance; PSO, pomegranate seed oil.

- Sco + Saline group: Received a single intraperitoneal (i.p.) injection of scopolamine at 3 mg/kg (19), followed by injections of 0.9% saline (1 mL/kg, i.p.) for 14 days.

- Sco + PSO (0.32 or 0.64 mg/kg) groups: Received a single i.p. injection of scopolamine at 3 mg/kg (19), followed by continuous injections of pomegranate seed oil at either 0.32 or 0.64 mg/kg (20) for 14 days.

- Saline + Sco group: Received continuous i.p. injections of saline for 14 days, followed by a single i.p. injection of scopolamine.

- The PSO (0.32 or 0.64 mg/kg) + Sco groups: Received continuous i.p. injections of pomegranate seed oil at either 0.32 or 0.64 mg/kg for 14 days, followed by a single i.p. injection of scopolamine (Figure 1).

Scopolamine hydrobromide was dissolved in 0.9% saline immediately before injection. The pomegranate seed oil was procured from Golnar Golestan Company (Gonbadkavos, Iran).

3.3. Inhibitory Passive Avoidance Memory Test

All rats were trained in a passive avoidance apparatus 24 hours before drug injection, and the memory retention test was conducted 24 hours after the final injection (19). The test spanned two days:

- Training day: Each rat was placed in the light chamber, and the latency time to enter the dark chamber was recorded (without foot shock). Rats that did not enter within a maximum of 2 minutes were excluded from the study. Thirty minutes later, the test

was repeated, during which the rat received a foot shock upon entering the dark chamber (50 Hz, 1 mA, 3 seconds). The rat was then promptly returned to its cage. After 2 minutes, the latency time was measured again (with a maximum of 120 seconds). Successful acquisition of inhibitory passive avoidance was determined for rats that did not enter the dark chamber within 2 minutes.

- Probe day: The step-through latency time was recorded for a maximum of 300 seconds.

3.4. Tissue Preparation

After anesthetization with ketamine (100 mg/mL) and xylazine (20 mg/mL), the rats were transcardially perfused with 4% paraformaldehyde (Scharlau, Spain). The brains were fixed for one week in 4% paraformaldehyde in PBS. Histological processing was performed using an automated tissue processor (Did Sabz, Iran), and the tissues were subsequently embedded in paraffin. The brains were sectioned in the coronal plane at 6 μ m (21) using a rotary microtome (Pooyan MK 1110, Iran). The sections were divided into two sets: One set was used for Congo red staining to detect A β plaques, and the other set was used for cresyl violet staining to visualize neurons.

3.5. Congo Red Staining

The sections were initially deparaffinized in xylene and rehydrated through a descending alcohol series. The brain sections were then stained by immersion in

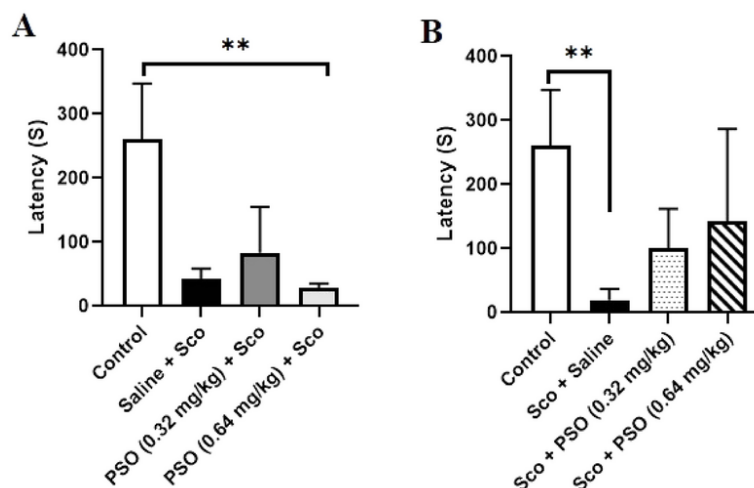


Figure 2. A and B, Passive avoidance test. Pomegranate seed oil improved the step-through latency time in treatment (B) group rats; however, it was not significant. B, the result showed that Sco injection in Sco + Saline could significantly decrease the memory, ($P < 0.01$). Data were expressed as the mean \pm SD. Kruskal-Wallis's test followed by Dunn's multiple comparisons test. ** $P < 0.01$. Abbreviations: PSO, pomegranate seed oil; Sco, scopolamine.

Congo red (DDK, Italy) for one hour at room temperature, followed by washing in distilled water. Subsequently, the sections were quickly differentiated with an alcoholic solution and washed for 5 minutes. For counterstaining, hematoxylin solution was applied for 5 minutes, followed by a 10-minute wash. The slides were dehydrated using 95% and 100% alcohols and cleared in xylene. Finally, all slides were coverslipped with Entellan glue (Merck, Germany) (22).

3.6. Cresyl Violet Staining

Brain sections were dewaxed in xylene, rehydrated through an alcohol series, and washed with distilled water. The sections were then stained in 0.02% cresyl violet (Sigma, USA) for 5 minutes. Following dehydration with 96% and 100% alcohols and clearing in xylene, the brain sections were coverslipped with Entellan glue (23).

3.7. Estimating the Number of Neurons and A β Plaques

Images were captured using an Olympus BX53 microscope at 40x magnification, equipped with a digital camera (DP73, Olympus, Japan). The CA1, CA3, and dentate gyrus (DG) subfields of the hippocampus were evaluated. Quantitative analyses of the images were conducted using cellSens Standard 1.14 software (Olympus, Tokyo, Japan). The number of neurons and A β

plaques in the hippocampus were randomly counted within a field of 12,000 μm^2 (24, 25).

3.8. Statistical Analysis

The results were presented as mean \pm SD. Since the data distribution was not normal, the Kruskal-Wallis's test was conducted, followed by Dunn's multiple comparisons test. Statistical significance was set at $P < 0.05$.

4. Results

4.1. Pomegranate Seed Oil Effects Step-Through Latency Time

The results showed that the Sco + Saline group exhibited a significant reduction in step-through latency time to enter the dark chamber compared to the control group (Figure 2 $P < 0.01$). In the PSO (0.32 mg/kg) + Sco and Sco + PSO (0.32 mg/kg) groups, the step-through latency increased relative to the saline groups; however, these increases were not statistically significant (Figure 2A and B). The Sco + PSO (0.64 mg/kg) group showed an improvement in latency time compared to the Sco + Saline group. The PSO (0.64 mg/kg) + Sco group did not demonstrate effectiveness in enhancing memory performance compared to the Saline + Sco group. Conversely, it significantly decreased latency time compared to the control group (Figure 2 $P < 0.01$).

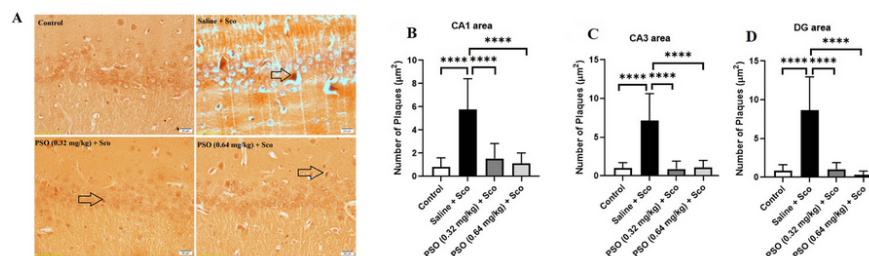


Figure 3. A - D, The density of hippocampal A β plaques in pretreatment groups. A, Congo red stained A β plaques were indicated by arrows ($\times 40$); B, pretreatment with pomegranate seed oil reduced the number of A β plaques in the rat hippocampal CA1; C, CA3; and D, DG areas. Data were expressed as the mean \pm SD. Kruskal-Wallis's test followed by Dunn's multiple comparisons test. **** $P < 0.0001$. The scale bar shows 20 μ m. Abbreviations: PSO, pomegranate seed oil; Sco, scopolamine.

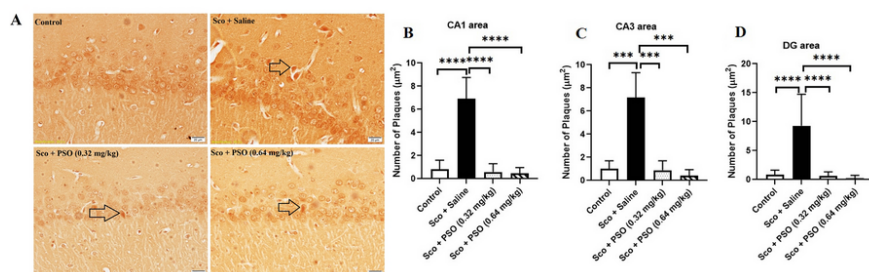


Figure 4. A - D, Hippocampal A β plaques density in treatment groups. A, Congo red stained A β plaques were indicated by arrows ($\times 40$); B, treatment with pomegranate seed oil reduced the number of A β plaques in the rat hippocampal CA1; C, CA3; and D, DG areas. Data were expressed as the mean \pm SD. Kruskal-Wallis's test followed by Dunn's multiple comparisons test. **** $P < 0.0001$, *** $P < 0.001$. The scale bar shows 20 μ m. Abbreviations: PSO, pomegranate seed oil; Sco, scopolamine.

4.2. Pomegranate Seed Oil Declined the Density of A β Plaques

Figure 3A and 4A demonstrated a notable increase in the hippocampal density of A β plaques in the Saline + Sco and Sco + Saline rats compared to the control group (Figure 3 - C and 4B - C, $P < 0.0001$). Treatment with PSO, both before and after the injection of scopolamine, significantly reduced the density of A β plaques across all areas of the hippocampus relative to the Sco + Saline and Saline + Sco groups (Figures 3 and 4, $P < 0.0001$). However, no significant differences were observed in the density of hippocampal A β plaques among the experimental groups receiving PSO.

4.3. Pomegranate Seed Oil Enhanced the Density of Neurons

Figures 5A and 6A illustrate the effect of pomegranate seed oil on the number of cresyl violet-

stained neurons before and after scopolamine administration. As shown in Figures 5B - D and Figure 6B - D, exposure to scopolamine significantly decreased the number of neurons ($P < 0.0001$) compared to the control group. Injection of pomegranate seed oil at doses of 0.32 and 0.64 mg/kg significantly increased the number of neurons ($P < 0.0001$) compared to the Sco + Saline and Saline + Sco groups.

5. Discussion

In agreement with other studies that used scopolamine to model AD-like conditions (26, 27), our findings confirmed impaired memory in the scopolamine-injected groups compared to the control group. We found that PSO re-established scopolamine-induced impairment in passive avoidance memory. Furthermore, we observed a notable reduction in the density of A β plaques and an increase in the number of neurons across various subfields of the hippocampus,

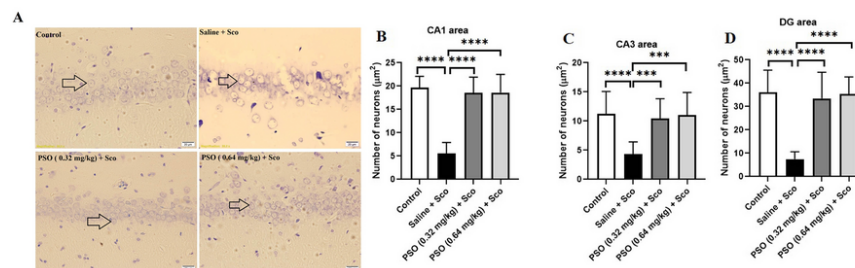


Figure 5. A - D, Hippocampal neuronal density in pretreatment groups. A, cresyl violet stained neurons were indicated by arrows ($\times 40$); B, pretreatment with pomegranate seed oil boosted the neuronal density in the rat hippocampal CA1; C, CA3; and D, DG areas. Data were expressed as the mean \pm SD. Kruskal-Wallis's test followed by Dunn's multiple comparisons test. **** $P < 0.0001$, *** $P < 0.001$. The scale bar shows 20 μm . Abbreviations: PSO, pomegranate seed oil; Sco, scopolamine.

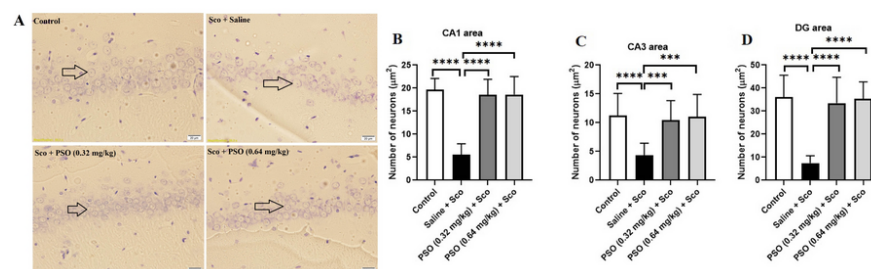


Figure 6. A - D, Hippocampal neuronal density in treatment groups. A, cresyl violet stained neurons were indicated by arrows ($\times 40$); B, treatment with pomegranate seed oil enhanced the neuronal density in the rat hippocampal CA1; C, CA3; and D, DG areas. Data were expressed as the mean \pm SD. Kruskal-Wallis's test followed by Dunn's multiple comparisons test. **** $P < 0.0001$, *** $P < 0.001$. The scale bar shows 20 μm . Abbreviations: PSO, pomegranate seed oil; Sco, scopolamine.

the main structure involved in memory formation and stabilization.

Pomegranate is recognized as a medicinal plant that offers effective neuroprotection against several neurodegenerative disorders (28-30). Sarkaki (15), in 2013, reported that continuous treatment with pomegranate seed extract for 14 days could ameliorate both passive and active memory impairments in cerebral ischemia model rats, due to the phytoestrogen and antioxidative effects of pomegranate seed extract (10). Another study revealed that pomegranate juice at a 20% concentration for 29 days improved learning and memory while protecting brain cells from degeneration induced by aluminum chloride (31). Moreover, pomegranate juice has been found to enhance spatial learning performance by reducing amyloid formation in AD model transgenic mice (7).

Dietary supplementation with pomegranate has shown promising effects on cognitive functions, particularly in APPsw/Tg2576 mice, where a 4%

pomegranate diet improved memory, learning, locomotor functions, and reduced anxiety (14). The results of the current study showed that treatment with 0.32 mg/kg and 0.64 mg/kg of pomegranate seed oil could improve passive avoidance memory in scopolamine-treated rats; however, these improvements were not significant. Remarkably, pretreatment with 0.64 mg/kg of PSO resulted in decreased memory performance.

The dysfunction of the cholinergic system results in the deterioration of learning and memory processing in AD. Scopolamine blocks acetylcholine muscarinic receptors, leading to increased acetylcholinesterase activity in the hippocampus and cortex (32). This enhancement is associated with memory deficits and oxidative stress in the brain. Notably, PSO has been shown to possess neuroprotective properties by inhibiting acetylcholinesterase activity and enhancing antioxidant capacity (33).

Synaptic plasticity is essential for the maintenance of learning and memory, which is impaired in AD pathology. Administration of 4% pomegranate over 15 months ameliorated the loss of key synaptic proteins, including PSD-95, Munc18-1, SNAP25, and synaptophysin (34). These proteins are vital for synaptic function and plasticity, indicating that pomegranate may play a significant role in enhancing cognitive functions through its protective effects on synaptic integrity.

The protective effects of PSO and pomegranate extract are thought to stem from their ability to neutralize reactive oxygen species (ROS), upregulate the expression of antioxidant genes, and exert anti-inflammatory, anti-apoptotic, and ATP-replenishing effects (11, 35). Studies have demonstrated that PSO protects against toxins such as mercuric chloride, hexachlorobutadiene, diazinon, cisplatin, gentamicin, and H₂O₂ (36, 37). In addition, pretreatment with pomegranate extract before ischemia/reperfusion in rats resulted in protection against brain injury and DNA damage (35).

Amyloid aggregation is a crucial pathological feature of AD (38). Deposition of A β peptides, especially A β 1-42, increases oxidative stress, neuroinflammation, and neuronal death (39). The administration of scopolamine increased A β production and oxidative stress while reducing neuronal density in rodent brains (19, 38). These findings align with the current study, which demonstrated that a single injection of scopolamine (3 mg/kg) elevated the density of A β plaques and reduced neuronal density in the hippocampus of rats.

Previous studies have shown that pomegranate juice and extracts exert neuroprotective effects against AD pathogenesis in several transgenic animal models, though the exact bioactive compounds have not yet been fully identified. Urolithins are suggested to mediate the neuroprotective effects of pomegranate against AD, as they inhibit A β fibrillation *in vitro* (40). It has been reported that pomegranate prevents amyloidogenesis in SK-N-SH cells stimulated with interleukin-1 β . It is assumed that pomegranate is a potent nutritional strategy to slow neurodegeneration in AD (41). Pomegranate peel extract (800 mg/kg/day) reduced the density of A β plaques in A β peptide-treated mice (28). In transgenic mice, pomegranate juice hindered the accumulation of soluble A β 1-42 and amyloid deposition in the hippocampus (7).

Moreover, the phenolic compounds in PSO have demonstrated significant neuroprotective effects. Shrivastava et al. (2023) developed a stable microemulsion incorporating PSO as an adjuvant for galantamine hydrobromide (GHBr). The optimized ratio of GHBr to

PSO showed promising outcomes, including reduced toxicity, enhanced antioxidant activity, and protection against A β -induced cell death. The study highlights PSO as a potential treatment to enhance the efficacy of anti-Alzheimer's therapies (12). Additionally, PSO can inhibit key enzymes, reduce ROS, prevent microglial activation, inhibit tau protein hyperphosphorylation, maintain synaptic plasticity, exhibit anti-inflammatory activity, and suppress beta-secretase-1 (BACE-1) (42).

These benefits are largely attributed to punicalic acid, the primary bioactive compound in PSO, which is an omega-5 isomer of conjugated α -linoleic acid. Punicalic acid possesses potent antioxidant and anti-inflammatory properties, reducing oxidative damage and inflammation by upregulating peroxisome proliferator-activated receptors. It further mitigates AD pathology by reducing beta-amyloid deposition and tau hyperphosphorylation through mechanisms such as enhanced GLUT4 protein expression and inhibition of calpain activation (43).

Pomegranate peel extract has been shown to decrease apoptosis and chromatolysis in the DG and CA3 areas of the hippocampus, reduce neurofibrillary tangles and senile plaques, and restore Nissl granules (44, 45). Consistent with these findings, the current study discovered that pretreatment and treatment with pomegranate seed oil (0.32 and 0.64 mg/kg) can reduce the density of A β plaques in scopolamine-injected rats. Additionally, it increased the density of neurons in the rat hippocampus. These results confirm the neuroprotective properties of pomegranate seed oil against the adverse effects of AD pathogenesis. Essa et al. (46) observed that long-term administration of pomegranate (for 15 months) decreased the levels of A β 1-40 and A β 1-42 in the brains of transgenic AD mice. In the current study, the protective effects of PSO were observed during a two-week treatment, which reduced the density of A β plaques in the hippocampus of scopolamine-injected rats. It has also been reported that pomegranate extract is more protective when administered as a pretreatment rather than as a therapeutic, as it reduces the histopathological features of AD (44). However, this study found that both pretreatment and treatment with pomegranate seed oil can be protective against the extension of A β plaques and the loss of neuronal density.

5.1. Conclusions

This study highlights the neuroprotective effects of PSO in a scopolamine-induced Alzheimer's disease rat model. The PSO reduced amyloid- β plaques, preserved neuronal density in the hippocampus, and restored

memory function. While further research is needed to elucidate detailed mechanisms and validate these findings in humans, this study suggests that PSO is a promising agent for AD treatment.

5.2. Limitation

This study had several limitations. First, while the neuroprotective effects of PSO were demonstrated, the specific bioactive compounds responsible and their mechanisms of action were not fully clarified. Additionally, the molecular pathways through which PSO reduces amyloid- β plaques and oxidative stress remain unclear. These aspects should be further explored in future studies. Moreover, it is recommended to focus on clinical trials to confirm the neuroprotective effects of PSO and evaluate its potential as a therapeutic agent for Alzheimer's disease.

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Footnotes

Authors' Contribution: M. J.: Supervision, conceptualization, resources, funding acquisition; V. K.: Project administration, methodology, resources; E. N.: Investigation, writing-original draft; F. Y.: Investigation; E. J.: Writing-original draft; G. B.: Formal analysis, writing-review and editing.

Conflict of Interests Statement: The authors declare no conflict of interest.

Data Availability: The dataset presented in the study is available on request from the corresponding author during submission or after publication.

Ethical Approval: The study was approved by the Research Ethics Committee of Golestan University of Medical Sciences, Gorgan, Iran (Agreement License No. IR.GOUMS.REC.1397.260).

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References

1. Mary A, Mancuso R, Heneka MT. Immune Activation in Alzheimer Disease. *Annu Rev Immunol.* 2024;**42**(1):585-613. [PubMed ID: 38424470]. <https://doi.org/10.1146/annurev-immunol-101921-035222>.
2. Dhakal S, Kushairi N, Phan CW, Adhikari B, Sabaratnam V, Macreadie I. Dietary Polyphenols: A Multifactorial Strategy to Target Alzheimer's Disease. *Int J Mol Sci.* 2019;**20**(20). [PubMed ID: 31615073]. [PubMed Central ID: PMC6834216]. <https://doi.org/10.3390/ijms20205090>.
3. Rai R, Kalar PL, Jat D, Mishra SK. Naringenin mitigates nanoparticulate-aluminium induced neuronal degeneration in brain cortex and hippocampus through downregulation of oxidative stress and neuroinflammation. *Neurochem Int.* 2024;**178**:105799. [PubMed ID: 38950625]. <https://doi.org/10.1016/j.neuint.2024.105799>.
4. Rao YL, Ganaraja B, Murlimanju BV, Joy T, Krishnamurthy A, Agrawal A. Hippocampus and its involvement in Alzheimer's disease: a review. *3 Biotech.* 2022;**12**(2):55. [PubMed ID: 35116217]. [PubMed Central ID: PMC8807768]. <https://doi.org/10.1007/s13205-022-03123-4>.
5. Chen X, Drew J, Berney W, Lei W. Neuroprotective Natural Products for Alzheimer's Disease. *Cells.* 2021;**10**(6). [PubMed ID: 34070275]. [PubMed Central ID: PMC8225186]. <https://doi.org/10.3390/cells10061309>.
6. Ullah A, Khan A, Ahmed S, Irfan HM, Hafiz AA, Jabeen K, et al. A review of pomegranate supplementation: A promising remedial avenue for Alzheimer's disease. *Heliyon.* 2023;**9**(11). e22483. [PubMed ID: 38074891]. [PubMed Central ID: PMC10700657]. <https://doi.org/10.1016/j.heliyon.2023.e22483>.
7. Hartman RE, Shah A, Fagan AM, Schwetye KE, Parsadanian M, Schulman RN, et al. Pomegranate juice decreases amyloid load and improves behavior in a mouse model of Alzheimer's disease. *Neurobiol Dis.* 2006;**24**(3):506-15. [PubMed ID: 17010630]. <https://doi.org/10.1016/j.nbd.2006.08.006>.
8. Kandyli P, Kokkinomagoulos E. Food Applications and Potential Health Benefits of Pomegranate and its Derivatives. *Foods.* 2020;**9**(2). [PubMed ID: 31979390]. [PubMed Central ID: PMC7074153]. <https://doi.org/10.3390/foods9020122>.
9. Boroushaki MT, Mollazadeh H, Afshari AR. Pomegranate seed oil: A comprehensive review on its therapeutic effects. *Int J Pharm Sci Rev Res.* 2016;**7**(2):430.
10. Sarkaki A, Farbood Y, Hashemi S, Rafiei Rad M. Pomegranate seed hydroalcoholic extract improves memory deficits in ovariectomized rats with permanent cerebral hypoperfusion /ischemia. *Avicenna J Phytomed.* 2015;**5**(1):43-55. [PubMed ID: 25767756]. [PubMed Central ID: PMC4352532].
11. Al-Sabahi BN, Fatope MO, Essa MM, Subash S, Al-Busafi SN, Al-Kusaibi FS, et al. Pomegranate seed oil: Effect on 3-nitropropionic acid-induced neurotoxicity in PC12 cells and elucidation of unsaturated fatty acids composition. *Nutr Neurosci.* 2017;**20**(1):40-8. [PubMed ID: 25238165]. <https://doi.org/10.1179/1476830514Y.00000000155>.
12. Shrivastava M, Khunt D, Shrivastava M, Misra M. Studies on pomegranate seed oil enriched galantamine hydrobromide microemulsion: formulation, in vitro antioxidant and neuroprotective potential. *Pharm Dev Technol.* 2023;**28**(2):153-63. [PubMed ID: 36662596]. <https://doi.org/10.1080/10837450.2023.2171433>.
13. Loizzo MR, Aiello F, Tenuta MC, Leporini M, Falco T, Tundis R. Pomegranate (*Punica granatum* L.). *Nonvitamin and Nonmineral Nutritional Supplements.* 2019. p. 467-72. <https://doi.org/10.1016/b978-0-12-812491-8.00062-x>.
14. Subash S, Braidly N, Essa MM, Zayana AB, Ragini V, Al-Adawi S, et al. Long-term (15 mo) dietary supplementation with pomegranates from Oman attenuates cognitive and behavioral deficits in a transgenic mice model of Alzheimer's disease. *Nutrition.*

- 2015;**31**(1):223-9. [PubMed ID: [25441596](#)]. <https://doi.org/10.1016/j.nut.2014.06.004>.
15. Sarkaki A, Rezaei M, Gharib Naseri M, Rafieirad M. Improving active and passive avoidance memories deficits due to permanent cerebral ischemia by pomegranate seed extract in female rats. *Malays J Med Sci*. 2013;**20**(2):25-34. [PubMed ID: [23983574](#)]. [PubMed Central ID: [PMC3743996](#)].
16. Chen WN, Yeong KY. Scopolamine, a Toxin-Induced Experimental Model, Used for Research in Alzheimer's Disease. *CNS Neurol Disord Drug Targets*. 2020;**19**(2):85-93. [PubMed ID: [32056532](#)]. <https://doi.org/10.2174/1871527319666200214104331>.
17. Muhammad T, Ali T, Ikram M, Khan A, Alam SI, Kim MO. Melatonin Rescue Oxidative Stress-Mediated Neuroinflammation/Neurodegeneration and Memory Impairment in Scopolamine-Induced Amnesia Mice Model. *J Neuroimmune Pharmacol*. 2019;**14**(2):278-94. [PubMed ID: [30478761](#)]. <https://doi.org/10.1007/s11481-018-9824-3>.
18. Saeed M, Naveed M, BiBi J, Kamboh AA, Arain MA, Shah QA, et al. The Promising Pharmacological Effects and Therapeutic/Medicinal Applications of Punica Granatum L. (Pomegranate) as a Functional Food in Humans and Animals. *Recent Pat Inflamm Allergy Drug Discov*. 2018;**12**(1):24-38. [PubMed ID: [29473532](#)]. <https://doi.org/10.2174/1872213X12666180221154713>.
19. Gorgani S, Jahanshahi M, Elyasi L. Taurine Prevents Passive Avoidance Memory Impairment, Accumulation of Amyloid- β Plaques, and Neuronal Loss in the Hippocampus of Scopolamine-Treated Rats. *Neurophysiology*. 2019;**51**(3):171-9. <https://doi.org/10.1007/s11062-019-09810-y>.
20. Boroushaki MT, Asadpour E, Sadeghnia HR, Dolati K. Effect of pomegranate seed oil against gentamicin-induced nephrotoxicity in rat. *J Food Sci Technol*. 2014;**51**(11):3510-4. [PubMed ID: [26396355](#)]. [PubMed Central ID: [PMC4571228](#)]. <https://doi.org/10.1007/s13197-012-0881-y>.
21. Paxinos G, Watson C. *The rat brain in stereotaxic coordinates: hard cover edition*. Elsevier; 2006.
22. Jahanshahi M, Nikmahzar E, Sayyahi A. Vitamin E therapy prevents the accumulation of congophilic amyloid plaques and neurofibrillary tangles in the hippocampus in a rat model of Alzheimer's disease. *Iran J Basic Med Sci*. 2020;**23**(1):86-92. [PubMed ID: [32395206](#)]. [PubMed Central ID: [PMC7206846](#)]. <https://doi.org/10.22038/IJBMS.2019.38165.9067>.
23. Nikmahzar E, Jahanshahi M, Ghaemi A, Naseri GR, Moharreri AR, Lotfinia AA. Hippocampal serotonin-2A receptor-immunoreactive neurons density increases after testosterone therapy in the gonadectomized male mice. *Anat Cell Biol*. 2016;**49**(4):259-72. [PubMed ID: [28127501](#)]. [PubMed Central ID: [PMC5266105](#)]. <https://doi.org/10.5115/acb.2016.49.4.259>.
24. Nikmahzar E, Jahanshahi M, Babakordi F. Ginkgo biloba Extract Decreases Scopolamine-Induced Congophilic Amyloid Plaques Accumulation in Male Rat's Brain. *Jundishapur Journal of Natural Pharmaceutical Products*. 2018;**13**(4). <https://doi.org/10.5812/jjnpp.69143>.
25. Nikmahzar E, Jahanshahi M, Elyasi L, Saeidi M, Babakordi F, Bahlakeh G. Human chorionic gonadotropin attenuates amyloid-beta plaques induced by streptozotocin in the rat brain by affecting cytochrome c-ir neuron density. *Iran J Basic Med Sci*. 2019;**22**(2):166-72. [PubMed ID: [30834082](#)]. [PubMed Central ID: [PMC6396995](#)]. <https://doi.org/10.22038/ijbms.2018.31412.7569>.
26. Liu Z, Qin G, Mana L, Dong Y, Huang S, Wang Y, et al. GAPT regulates cholinergic dysfunction and oxidative stress in the brains of learning and memory impairment mice induced by scopolamine. *Brain Behav*. 2020;**10**(5). e01602. [PubMed ID: [32174034](#)]. [PubMed Central ID: [PMC7218254](#)]. <https://doi.org/10.1002/brb3.1602>.
27. Sayyahi A, Jahanshahi M, Amini H, Sepehri H. Vitamin E can compensate the density of M1 receptors in the hippocampus of scopolamine-treated rats. *Folia Neuropathol*. 2018;**56**(3):215-28. [PubMed ID: [30509043](#)]. <https://doi.org/10.5114/fn.2018.78703>.
28. Morzelle MC, Salgado JM, Telles M, Mourelle D, Bachiega P, Buck HS, et al. Neuroprotective Effects of Pomegranate Peel Extract after Chronic Infusion with Amyloid-beta Peptide in Mice. *PLoS One*. 2016;**11**(11). e0166123. [PubMed ID: [27829013](#)]. [PubMed Central ID: [PMC5102433](#)]. <https://doi.org/10.1371/journal.pone.0166123>.
29. Doostkam A, Bassiri-Jahromi S, Iravani K. Punica Granatum with Multiple Effects in Chronic Diseases. *International Journal of Fruit Science*. 2019;**20**(3):471-94. <https://doi.org/10.1080/15538362.2019.1653809>.
30. Nambiar T. The Effect of Ayurveda Antioxidants on Lifespan and Motor Skills in the Huntington's model of *Drosophila melanogaster*. *South Carolina Junior Academy of Sci*. 2023.
31. Mansour Suliman SA. Possible Neuroprotective Role of Pomegranate Juice in Aluminum Chloride Induced Alzheimer's Like Disease in Mice. *Journal of Alzheimer's Disease & Parkinsonism*. 2015;**5**(1). <https://doi.org/10.4172/2161-0460.1000188>.
32. Sun K, Bai Y, Zhao R, Guo Z, Su X, Li P, et al. Neuroprotective effects of matrine on scopolamine-induced amnesia via inhibition of AChE/BuChE and oxidative stress. *Metab Brain Dis*. 2019;**34**(1):173-81. [PubMed ID: [30406376](#)]. <https://doi.org/10.1007/s11011-018-0335-y>.
33. Amri Z, Ghorbel A, Turki M, Akroun FM, Ayadi F, Elfeki A, et al. Effect of pomegranate extracts on brain antioxidant markers and cholinesterase activity in high fat-high fructose diet induced obesity in rat model. *BMC Complement Altern Med*. 2017;**17**(1):339. [PubMed ID: [28655305](#)]. [PubMed Central ID: [PMC5488477](#)]. <https://doi.org/10.1186/s12906-017-1842-9>.
34. Braidy N, Essa MM, Poljak A, Selvaraju S, Al-Adawi S, Manivasagam T, et al. Consumption of pomegranates improves synaptic function in a transgenic mice model of Alzheimer's disease. *Oncotarget*. 2016;**7**(40):64589-604. [PubMed ID: [27486879](#)]. [PubMed Central ID: [PMC5323101](#)]. <https://doi.org/10.18632/oncotarget.10905>.
35. Ahmed MA, El Morsy EM, Ahmed AA. Pomegranate extract protects against cerebral ischemia/reperfusion injury and preserves brain DNA integrity in rats. *Life Sci*. 2014;**110**(2):61-9. [PubMed ID: [25010842](#)]. <https://doi.org/10.1016/j.lfs.2014.06.023>.
36. Boroushaki MT, Rajabian A, Farzadnia M, Hoseini A, Poorlaskari M, Taghavi A, et al. Protective effect of pomegranate seed oil against cisplatin-induced nephrotoxicity in rat. *Ren Fail*. 2015;**1**:6. [PubMed ID: [26275112](#)].
37. Bihanta M, Hosseini A, Ghorbani A, Boroushaki MT. Protective effect of pomegranate seed oil against H(2)O(2)-induced oxidative stress in cardiomyocytes. *Avicenna J Phytomed*. 2017;**7**(1):46-53. [PubMed ID: [28265546](#)]. [PubMed Central ID: [PMC5329176](#)].
38. Hernandez-Rodriguez M, Arciniega-Martinez IM, Garcia-Marin ID, Correa-Basurto J, Rosales-Hernandez MC. Chronic Administration of Scopolamine Increased GSK3 β 9, Beta Secretase, Amyloid Beta, and Oxidative Stress in the Hippocampus of Wistar Rats. *Mol Neurobiol*. 2020;**57**(9):3979-88. [PubMed ID: [32638218](#)]. <https://doi.org/10.1007/s12035-020-02009-x>.
39. Fanlo-Ucar H, Picon-Pages P, Herrera-Fernandez V, Ill-Raga G, Munoz FJ. The Dual Role of Amyloid Beta-Peptide in Oxidative Stress and Inflammation: Unveiling Their Connections in Alzheimer's Disease Etiopathology. *Antioxidants (Basel)*. 2024;**13**(10). [PubMed ID: [39456461](#)]. [PubMed Central ID: [PMC11505517](#)]. <https://doi.org/10.3390/antiox13101208>.
40. Yuan T, Ma H, Liu W, Niesen DB, Shah N, Crews R, et al. Pomegranate's Neuroprotective Effects against Alzheimer's Disease Are Mediated by Urolithins, Its Ellagitannin-Gut Microbial Derived Metabolites. *ACS Chem Neurosci*. 2016;**7**(1):26-33. [PubMed ID: [26559394](#)]. <https://doi.org/10.1021/acschemneuro.5b00260>.

41. Velagapudi R, Baco G, Khela S, Okorji U, Olajide O. Pomegranate inhibits neuroinflammation and amyloidogenesis in IL-1 β -stimulated SK-N-SH cells. *Eur J Nutr*. 2016;**55**(4):1653-60. [PubMed ID: 26155780]. <https://doi.org/10.1007/s00394-015-0984-0>.
42. George N, AbuKhader M, Al Balushi K, Al Sabahi B, Khan SA. An insight into the neuroprotective effects and molecular targets of pomegranate (*Punica granatum*) against Alzheimer's disease. *Nutr Neurosci*. 2023;**26**(10):975-96. [PubMed ID: 36125072]. <https://doi.org/10.1080/1028415X.2022.2121092>.
43. Guerra-Vazquez CM, Martinez-Avila M, Guajardo-Flores D, Antunes-Ricardo M. Punicic Acid and Its Role in the Prevention of Neurological Disorders: A Review. *Foods*. 2022;**11**(3). [PubMed ID: 35159404]. [PubMed Central ID: PMC8834450]. <https://doi.org/10.3390/foods11030252>.
44. Almuhayawi MS, Ramadan WS, Harakeh S, Al Jaouni SK, Bharali DJ, Mousa SA, et al. The potential role of pomegranate and its nano-formulations on cerebral neurons in aluminum chloride induced Alzheimer rat model. *Saudi J Biol Sci*. 2020;**27**(7):1710-6. [PubMed ID: 32565686]. [PubMed Central ID: PMC7296487]. <https://doi.org/10.1016/j.sjbs.2020.04.045>.
45. Hakeem K, Harakeh S, Ramadan W, Al Muhayawi M, Al Jaouni S, Mousa S. Pomegranate peel extract lessens histopathologic changes and restores antioxidant homeostasis in the hippocampus of rats with aluminium chloride-induced Alzheimer's disease. *Asian Pacific Journal of Tropical Medicine*. 2020;**13**(10). <https://doi.org/10.4103/1995-7645.291039>.
46. Essa MM, Subash S, Akbar M, Al-Adawi S, Guillemin GJ. Long-term dietary supplementation of pomegranates, figs and dates alleviate neuroinflammation in a transgenic mouse model of Alzheimer's disease. *PLoS One*. 2015;**10**(3). e0120964. [PubMed ID: 25807081]. [PubMed Central ID: PMC4373715]. <https://doi.org/10.1371/journal.pone.0120964>.